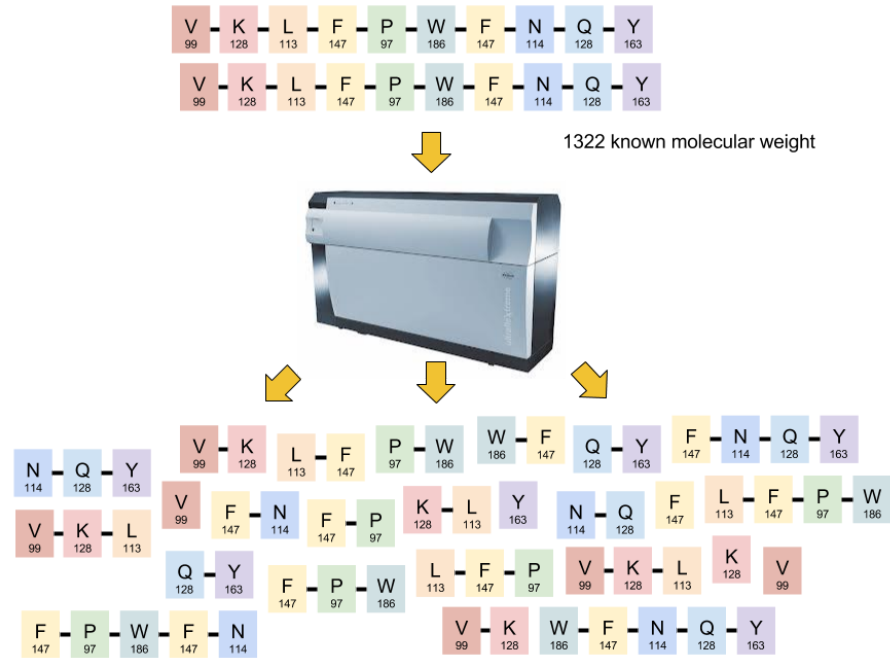


Determining a Peptide's Sequence

- From last time we learned that we can't always use DNA to resolve peptide/protein sequences
- What else can we do?
 - Extract and purify a pure sample of the peptide/protein
 - Try to resolve the peptide sequence by analyzing this sample
- Today's approach
 - Randomly fracture the peptide
 - Assemble an answer from the peices

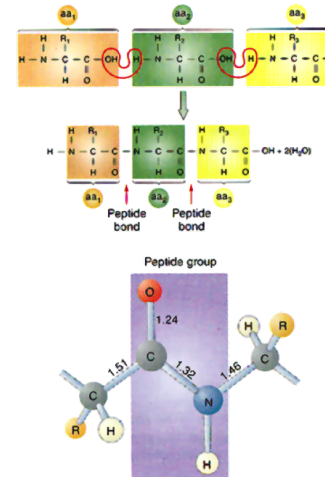


Molecular Weights are the Puzzle Peices



Structure of a Peptide Chain

- Peptides are chains of amino acids that are joined by *peptide* bonds
- These bonds reduce the weight of each amino acid by one H₂O molecule
- The result is called a *residue*
- A Mass Spectrograph can precisely measure the molecular weight (and charge and abundance) of any peptide chain
- Since the molecular weight of each of the possible 20 residues is known precisely, one can ask the question, which combination of residues would give a particular weight?
- The problem is ambiguous for the entire molecule
 - Consider all permutations of 'PIT':
'PIT', 'PTI', 'ITP', 'IPT', 'TPI', and 'TIP' all weigh the same
 - But they differ in their 2-peptide fragments:
'PIT' breaks into 'PI' and 'IT', while
'PTI' breaks into 'PT' and 'TI'



An Simplified Peptide Weight table

- The actual molecular weight of an amino acid is a real number. This accounts for the relative abundances of atomic isotopes
- Today, we will use a simplified version that assumes only integer molecular weights
- Example:

- Molecular weight of Glycine Amino Acid

$$W(C_2H_5NO_2) = 12 \times 2 + 5 \times 1 + 14 + 16 \times 2 = 75$$

- Molecular weight of Glycine Residue (Minus the H_2O lost forming the peptide bond)

$$W(C_2H_5NO_2 - H_2O) = 57$$

- We can repeat this for all 20 Amino Acids to get a integer molecular weight table, which I call *Daltons*

Table Definitions

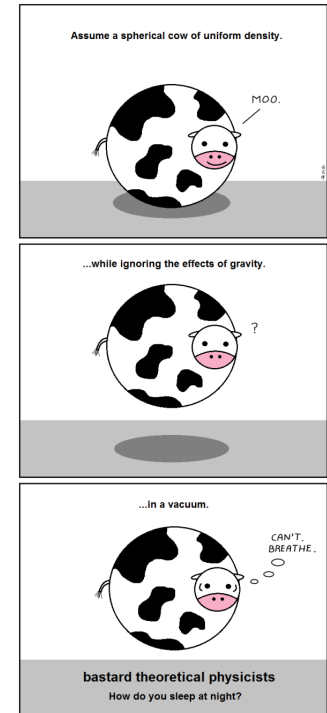
```
AminoAcid = {
  'A': 'Alanine', 'C': 'Cysteine', 'D': 'Aspartic acid', 'E': 'Glutamic acid',
  'F': 'Phenylalanine', 'G': 'Glycine', 'H': 'Histidine', 'I': 'Isoleucine',
  'K': 'Lysine', 'L': 'Leucine', 'M': 'Methionine', 'N': 'Asparagine',
  'P': 'Proline', 'Q': 'Glutamine', 'R': 'Arginine', 'S': 'Serine',
  'T': 'Theronine', 'V': 'Valine', 'W': 'Tryptophan', 'Y': 'Tyrosine',
  '*': 'STOP'
}

AminoAbbrv = {
  'A': 'Ala', 'C': 'Cys', 'D': 'Asp', 'E': 'Glu',
  'F': 'Phe', 'G': 'Gly', 'H': 'His', 'I': 'Ile',
  'K': 'Lys', 'L': 'Leu', 'M': 'Met', 'N': 'Asn',
  'P': 'Pro', 'Q': 'Gln', 'R': 'Arg', 'S': 'Ser',
  'T': 'Thr', 'V': 'Val', 'W': 'Trp', 'Y': 'Tyr',
  '*': 'STP'
}

# Here's a new dictionary!
Daltons = {
  'A': 71, 'C': 103, 'D': 115, 'E': 129,
  'F': 147, 'G': 57, 'H': 137, 'I': 113,
  'K': 128, 'L': 113, 'M': 131, 'N': 114,
  'P': 97, 'Q': 128, 'R': 156, 'S': 87,
  'T': 101, 'V': 99, 'W': 186, 'Y': 163
}
```

Some Issues with our Table

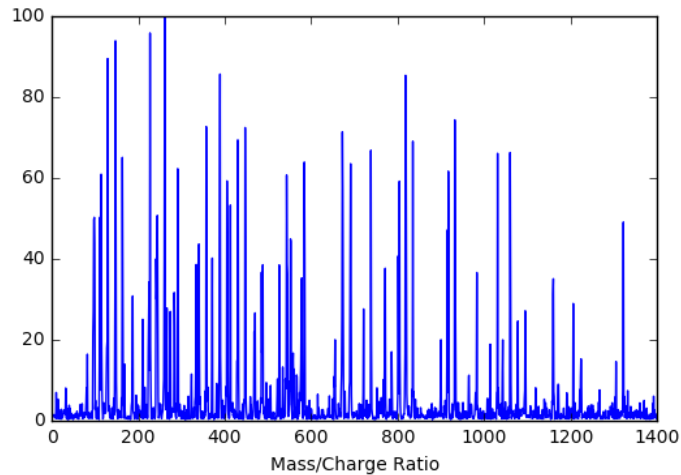
- We can't distinguish between Leucine (L) and Isoleucine (I). They both weight 113 d
- Nor can we distinguish Lysine (K) and Glutamine (Q), which weigh 128 d
- For long peptide chains >50, our errors can build up
- In reality, peptides can loose or gain one or more small molecules from their side chains and fractured peptide bonds
 - Gain Hydrogen ions (H, +1 Dalton)
 - Lose Water (H₂O, -18 Daltons)
 - Lose Ammonia (NH₃, -17 Daltons)
- This leads to measurements that vary around the ideal sums we assume
- Regardless of these caveats, let's keep going



The total molecular weight of our target

```
TyrocidineB1 = "VKLFPWFNQY"  
  
# The weight of Tyrocidine B1  
print sum([Daltons[res] for res in TyrocidineB1])
```

1322



- Generally, we will assume that the peptide's total molecular weight is known
- We will use it as a terminating condition for many of our algorithms that attempt to reconstruct the measured set of weights

Ideally, what Weights should we get?

- We will make the optimistic assumption that we will fracture our given peptide chain into all of its constituent parts
- For a 10 peptide chain
 - 10 single peptides
 - 9, 2-peptide chains
 - 8, 3-peptide chains
 - 7, 4-peptide chains
 - 6, 5-peptide chains
 - 5, 6-peptide chains
 - 4, 7-peptide chains
 - 3, 8-peptide chains
 - 2, 9-peptide chains
 - 1, 10-peptide chain
- This gives an upper bound of $\binom{11}{2} = 55$ molecular weights
- In reality both the peptide chains and their weights may not be unique
- The collection of all possible sub-peptide molecular weights from a peptide is called the peptide's *Theoretical Spectrum*

Code for computing a Theoretical Spectrum

```
def TheoreticalSpectrum(peptide):  
    # Generate every possible fragment of a peptide  
    spectrum = set()  
    for fragLength in xrange(1, len(peptide)+1):  
        for start in xrange(0, len(peptide)-fragLength+1):  
            seq = peptide[start:start+fragLength]  
            spectrum.add(sum([Daltons[res] for res in seq]))  
    return sorted(spectrum)  
  
print TyrocidineB1  
spectrum = TheoreticalSpectrum(TyrocidineB1)  
print len(spectrum)  
print spectrum
```

VKLFQWFNQY

51

[97, 99, 113, 114, 128, 147, 163, 186, 227, 241, 242, 244, 260, 261, 283, 291, 333, 340, 357, 388, 389, 405, 430, 447, 485, 487, 543, 544, 552, 575, 577, 671, 672, 690, 691, 738, 770, 804, 818, 819, 835, 917, 932, 982, 1031, 1060, 1095, 1159, 1223, 1322]

- Why are we using a set rather than a list? Notice that we end up returning a list.

Fragments and their Spectrums

```
peptide = TyrocidineB1
fragList = []
for fragLength in xrange(1, len(peptide)+1):
    for start in xrange(0, len(peptide)-fragLength+1):
        seq = peptide[start:start+fragLength]
        fragList.append((sum([Daltons[res] for res in seq]), seq))

print peptide
print len(fragList)
N = 0
lastWeight = 0
for weight, frag in sorted(fragList):
    print "%12s: %4d%s" % (frag, weight, "*" if (weight == lastWeight) else " "),
    N += 1
    if (N % 5 == 0):
        print
        lastWeight = weight
```

VKLFPWFNQY

55

P: 97	V: 99	L: 113	N: 114	K: 128
Q: 128*	F: 147	F: 147*	Y: 163	W: 186
VK: 227	KL: 241	NQ: 242	FP: 244	LF: 260
FN: 261	PW: 283	QY: 291	WF: 333	VKL: 340
LFP: 357	KLF: 388	FNQ: 389	NQY: 405	FPW: 430
PWF: 430*	WFN: 447	KLFP: 485	VKLF: 487	LFPW: 543
PWFN: 544	FNQY: 552	WFNQ: 575	FPWF: 577	VKLFP: 584
KLFPW: 671	PWFNQ: 672	LFPWF: 690	FPWFN: 691	WFNQY: 738
VKLFPW: 770	LFPWFN: 804	KLFPWF: 818	FPWFNQ: 819	PWFNQY: 835
VKLFPWF: 917	KLFPWFN: 932	LFPWFNQ: 932*	FPWFNQY: 982	VKLFPWFN: 1031
KLFPWFNQ: 1060	LFPWFNQY: 1095	VKLFPWFNQ: 1159	KLFPWFNQY: 1223	VKLFPWFNQY: 1322

Let's try a smaller example

```
peptide = 'PLAY'
spectrum = TheoreticalSpectrum(peptide)
print len(spectrum), spectrum

fragList = []
for fragLength in xrange(1, len(peptide)+1):
    for start in xrange(0, len(peptide)-fragLength+1):
        seq = peptide[start:start+fragLength]
        fragList.append((sum([Daltons[res] for res in seq]), seq))

print len(fragList)
N = 0
lastWeight = 0
for weight, frag in sorted(fragList):
    print "%12s: %4d%s" % (frag, weight, "*" if (weight == lastWeight) else " "),
    N += 1
    if (N % 5 == 0):
        print
        lastWeight = weight
```

```
10 [71, 97, 113, 163, 184, 210, 234, 281, 347, 444]
10
      A:  71          P:  97          L: 113          Y: 163          LA: 184
      PL: 210        AY: 234        PLA: 281        LAY: 347        PLAY: 444
```

Can we Invert the Process of creating a Spectrum?

- In essence, the problem of inferring a peptide chain from the set of mass values reported by a Mass Spectrometer is the inverse of the code we just wrote

Easy Problem: Peptide Sequence \rightarrow Spectrum

Hard Problem: Peptide Sequence \leftarrow Spectrum

- Why is computing a spectrum from a peptide sequence easy? $O(N^2)$?
- Why is computing a peptide sequence from a spectrum hard? $O(?)$

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"I'm trying to back it up, but I can't find reverse"

How might you approach this problem?

- Can you think of a Brute-Force way of solving this problem?
- Here's one:
 1. For every peptide sequence with the target peptide's molecular weight
 2. Compute the sequence's Theoretical Spectrum
 3. If it matches the one given, report this peptide as a possible solution
- Which step in this algorithm is the hard part?



A Brute-Force Attempt

```
def PossiblePeptide(spectrum, prefix=''):
    """ A brute force method of generating all peptide sequences that add up to our target weight from the given spectrum """
    global peptideList
    if (len(prefix) == 0):
        peptideList = []
    current = sum([Daltons[res] for res in prefix])
    target = max(spectrum) # our target
    if (current == target):
        peptideList.append(prefix)
    elif (current < target):
        for residue in Daltons.iterkeys():
            PossiblePeptide(spectrum, prefix+residue)

def TestPeptides(candidateList, target):
    filteredList = []
    for peptide in candidateList:
        candidateSpectrum = TheoreticalSpectrum(peptide)
        if (candidateSpectrum == target):
            filteredList.append(peptide)
    return filteredList

spectrum = TheoreticalSpectrum('PLAY')
%time PossiblePeptide(spectrum)
print len(peptideList), "candidates", "PLAY" in peptideList
%time matches = TestPeptides(peptideList, spectrum)
print matches, "PLAY" in matches
```

```
CPU times: user 3.57 s, sys: 3 ms, total: 3.58 s
Wall time: 3.57 s
3687 candidates True
CPU times: user 73 ms, sys: 0 ns, total: 73 ms
Wall time: 73.2 ms
['PIAY', 'PLAY', 'YAIP', 'YALP'] True
```

Impressions?

- Not so bad for a first attempt, but how will it perform for longer peptides?
- We are getting the expected answer as well as answers with the indistinguishable amino acids substituted
- We are also getting the sequence reversed? Is this a surprise?
- We could code around this, but for today we'll just include the reversed peptide chain as a possible answer

Could we do better?

- The brute force method does not make good use of the spectrum it is given
- It only ever considers the largest value from this table
- How might we make use of the other values?

Improving on Brute Force

- We could extend our prefix using *only* residues that appear in our spectrum
- The weight of every new prefix that we consider should also be in our spectrum

Actual fragments: P L A Y PL LA AY PLA LAY PLAY

Growing and Checking prefixes:

A	I	L	P	Y
AI = LA	IA = LA	LA = LA	PI = PL	YA = AY
AIP = PLA	IAP = PLA	LAP = PLA	PIA = PLA	YAI = LAY
AIPY = PLAY	IAPY = PLAY	LAPY = PLAY	PIAY = PLAY	YAIP = PLAY
AIY = LAY	IAY = LAY	LAY = LAY		YAL = LAY
AIYP = PLAY	IAYP = PLAY	LAYP = PLAY		YALP = PLAY
AL = LA	IP = PL	LP = PL	PL = PL	
ALP = PLA	IPA = PLA	LPA = PLA	PLA = PLA	
ALPY = PLAY	IPAY = PLAY	LPAY = PLAY	PLAY = PLAY	
ALY = LAY				
ALYP = PLAY				
AY = AY				
AYI = LAY				
AYIP = PLAY				
AYL = LAY				
AYLP = PLAY				

Only a Small Change to the Code

```
def ImprovedPossiblePeptide(spectrum, prefix=''):
    global peptideList
    if (len(prefix) == 0):
        peptideList = []
    current = sum([Daltons[res] for res in prefix])
    target = max(spectrum)
    if (current == target):
        peptideList.append(prefix)
    elif (current < target):
        for residue in Daltons.iterkeys():
            # make sure that this residue appears in our spectrum
            if (Daltons[residue] not in spectrum):
                continue
            # make sure that adding this residue to the sequence we have so far appears in our spectrum
            extend = prefix + residue
            if (sum([Daltons[res] for res in extend]) not in spectrum):
                continue
            ImprovedPossiblePeptide(spectrum, extend)

spectrum = TheoreticalSpectrum('PLAY')
%time ImprovedPossiblePeptide(spectrum)
print len(peptideList), "PLAY" in peptideList
print peptideList
%time matches = TestPeptides(peptideList, spectrum)
print matches, "PLAY" in matches

CPU times: user 1 ms, sys: 0 ns, total: 1 ms
Wall time: 761 µs
16 True
['AIPY', 'AIYP', 'ALPY', 'ALYP', 'AYIP', 'AYLP', 'IAPY', 'IAYP', 'IPAY', 'LAPY', 'LAYP', 'LPAY', 'PIAY', 'PLAY', 'YAIP', 'YALP']
CPU times: user 0 ns, sys: 0 ns, total: 0 ns
Wall time: 404 µs
['PIAY', 'PLAY', 'YAIP', 'YALP'] True
```

Impact of a small change

- Provides a HUGE performance difference
- Yet another example of Branch-and-Bound
- We improved both the enumeration and verification phases, but the difference was much more significant in the enumeration step

```
for peptide in peptideList:  
    print peptide,
```

```
AIPY AIYP ALPY ALYP AYIP AYLP IAPY IAYP IPAY LAPY LAYP LPAY PIAY PLAY YAIP YALP
```

```
TheoreticalSpectrum('PLAY')
```

```
[71, 97, 113, 163, 184, 210, 234, 281, 347, 444]
```

```
TheoreticalSpectrum('LAPY')
```

```
[71, 97, 113, 163, 168, 184, 260, 281, 331, 444]
```

```
print sum([Daltons[res] for res in 'AP']) # Suffix of 'LAP' prefix  
print sum([Daltons[res] for res in 'APY']) # Suffix of 'LAPY'  
print sum([Daltons[res] for res in 'PY']) # Suffix of 'LAPY'
```

```
168  
331  
260
```

- There are still differences in the spectrums, yet every prefix was in the spectrum when we added it. What are we missing?
- Suffixes!

We can do Even Better

- All *suffixes* of each prefix that we consider should also be in our spectrum

```
def UltimatePossiblePeptide(spectrum, prefix=''):
    global peptideList
    if (len(prefix) == 0):
        peptideList = []
    current = sum([Daltons[res] for res in prefix])
    target = max(spectrum)
    if (current == target):
        peptideList.append(prefix)
    elif (current < target):
        for residue in Daltons.iterkeys():
            extend = prefix + residue
            # test every new suffix created by adding this new residue
            # Note: this includes the residue itself as the length 1 suffix
            suffix = [extend[i:] for i in xrange(len(extend))]
            for fragment in suffix:
                if (sum([Daltons[res] for res in fragment]) not in spectrum):
                    break
            else:
                UltimatePossiblePeptide(spectrum, extend)

spectrum = TheoreticalSpectrum('PLAY')
%time UltimatePossiblePeptide(spectrum)
print len(peptideList), peptideList, "PLAY" in peptideList
%time matches = TestPeptides(peptideList, spectrum)
print matches, "PLAY" in matches
```

```
CPU times: user 3 ms, sys: 0 ns, total: 3 ms
Wall time: 2.44 ms
4 ['PIAY', 'PLAY', 'YAIP', 'YALP'] True
CPU times: user 0 ns, sys: 0 ns, total: 0 ns
Wall time: 147 µs
['PIAY', 'PLAY', 'YAIP', 'YALP'] True
```

- A little slower, but our list is pruned significantly
- All of these have identical spectrums

Now let's return to our *real* peptide

```
spectrum = TheoreticalSpectrum(TyrocidineB1)
%time UltimatePossiblePeptide(spectrum)
print len(peptideList)
print TyrocidineB1 in peptideList
%time matches = TestPeptides(peptideList, spectrum)
print len(matches)
print TyrocidineB1 in matches
```

```
CPU times: user 66 ms, sys: 8 ms, total: 74 ms
Wall time: 63.9 ms
16
True
CPU times: user 1e+03 µs, sys: 0 ns, total: 1e+03 µs
Wall time: 1.55 ms
16
True
```

```
print TyrocidineB1
for i, peptide in enumerate(peptideList):
    print peptide,
    if (i % 4 == 3):
        print
```

```
VKLFPFNQY
VKIFPWFNKY VKIFPWFNQY VKLFPWFNKY VKLFPWFNQY
VQIFPWFNKY VQIFPWFNQY VQLFPWFNKY VQLFPWFNQY
YKNFWPFIKV YKNFWPFIQV YKNFWPFLKV YKNFWPFLQV
YQNFWPFIKV YQNFWPFIQV YQNFWPFLKV YQNFWPFLQV
```

All of these peptides give also give us our desired spectrum

Great, but our assumptions are a little Naïve

- In reality, Mass Spectrometers don't report the Theoretical Spectrum of a peptide
- Instead they report a measured or *Experimental Spectrum*
- This spectrum might *miss* some fragments
- It might also report *false* fragments
 - From Contaminants
 - New peptides formed by unintended reactions between fragments
- The result is that some of the masses that appear may be misleading, and some that we want might be missing
- We need to develop algorithms for reporting candidate protein sequences that are robust to noise

Example experimental spectrum for Tyrocidine B1

97,	99,	113,	114,	128,	147,	163,
186,	200,	227,	241,	242,	244,	260,
261,	283,	291,	333,	340,	357,	388,
389,	405,	430,	447,	457,	485,	487,
543,	544,	552,	575,	577,	584,	659,
671,	672,	690,	691,	731,	738,	770,
804,	818,	819,	835,	906,	917,	932,
982,	1031,	1060,	1095,	1159,	1223,	1322

False Masses: present in the experimental spectrum, but not in the theoretical spectrum

Missing Masses: present in the theoretical spectrum, but not in the experimental spectrum

Example experimental spectrum for Tyrocidine B1

97,	99,	113,		128,	147,	163,
186,	200 ,	227,	241,	242,	244,	260,
261,	283,	291,	333,	340,	357,	
	405,	430,	447,	457 ,		487,
543,	544,	552,	575,	577,	584,	659 ,
671,	672,	690,	691,	731 ,	738,	770,
804,	818,	819,	835,	906 ,	917,	932,
982,	1031,		1095,	1159,		1322

False Masses: We don't know which these are

Missing Masses: And these values don't appear

An aside: Faking an Experimental Spectrum

```
# generate a synthetic experimental spectrum with 10% Error
import itertools
import random
random.seed(1961)

spectrum = TheoreticalSpectrum(TyrocidineB1)

# Pick around ~10% at random to remove
missingMass = random.sample(spectrum[:-1], 6) # keep largest mass
print "Missing Masses = ", missingMass

# Add back another ~10% of false, but actual, peptide masses
falseMass = []
for i in xrange(5):
    fragment = ''.join(random.sample(Daltons.keys(), random.randint(2, len(TyrocidineB1)-2)))
    weight = sum([Daltons[residue] for residue in fragment])
    falseMass.append(weight)
print "False Masses = ", falseMass

experimentalSpectrum = sorted(set([mass for mass in spectrum if mass not in missingMass] + falseMass))

Missing Masses = [1159, 114, 691, 186, 819, 357]
False Masses = [457, 200, 731, 906, 659]
```

```
print experimentalSpectrum
```

```
[97, 99, 113, 128, 147, 163, 200, 227, 241, 242, 244, 260, 261, 283, 291, 333, 340, 388, 389, 405, 430, 447, 457, 485, 487, 543, 544, 552, 575, 577, 584, 659, 671, 672, 690, 731, 738, 770, 804, 818, 835, 906, 917, 932, 982, 1031, 1060, 1095, 1223, 1322]
```


A Golf Tournament Analogy

- After the first couple of rounds of a major golf tournament a *cut* is made of all golfers who are so far back from the leader that it is deemed they are unlikely to ever finish in the money
- These *cut* golfers are removed from further consideration
- This choice is *heuristic*
 - It is possible that a player just below the cut could have two exceptional rounds, but that is considered unlikely
- What is the equivalent of a score in our peptide finding problem?
 - The number of matching masses in the candidate peptide's Theoretical Spectrum and the Experimental Spectrum
 - Normalized score, why?
 - $\text{len}(\text{intersection of candidate and experimental spectrums}) / \text{len}(\text{union of candidate and experimental spectrums})$
 - **Jaccard Index** for sets
- In our peptide *golf game* a round will be considered a one peptide extension of a active set of *player* peptides
- We will do cuts on every round, keeping to top 5% of finishers or the top 5 players, which ever is more
- Why 5%? It is arbitrary, but on each round we will extend the current set of players by one of 20 amino acids, thus increasing the number of peptides by a factor of 20, so reducing by 5% leaves the poolsize relatively stable.

POS	CTRY	PLAYER	TO PAR	R1	R2	R3	R4	TOT
1		Webb Simpson	+1	72	73	68	68	281
T2		Michael Thompson	+2	66	75	74	67	282
T2		Graeme McDowell	+2	69	72	68	73	282
T4		Jason Dufner	+3	72	71	70	70	283
T4		Padraig Harrington	+3	74	70	71	68	283
T4		David Toms	+3	69	70	76	68	283
T4		John Peterson	+3	71	70	72	70	283
T4		Jim Furyk	+3	70	69	70	74	283
9		Ernie Els	+4	75	69	68	72	284
T10		John Senden	+5	72	73	68	72	285
T10		Kevin Chappell	+5	74	71	68	72	285
T10		Casey Wittenberg	+5	71	77	67	70	285
T10		Rebec Goosen	+5	75	70	69	71	285
T10		Lee Westwood	+5	73	72	67	73	285
T15		Martin Kaymer	+6	74	71	69	72	286
T15		Aaron Watkins	+6	72	71	72	71	286
T15		Fredrik Jacobson	+6	72	71	68	75	286
T15		Adam Scott	+6	76	70	70	286	

An Implementation

```
def LeaderboardFindPeptide(noisySpectrum, cutThreshold=0.05):
    # Golf Tournament Heuristic
    spectrum = set(noisySpectrum)
    target = max(noisySpectrum)
    players = [''.join(peptide) for peptide in itertools.product(Daltons.keys(), repeat=2)]
    round = 1
    currentLeader = [0.0, '']
    while True:
        print "%8d Players in round %d [%5.4f]" % (len(players), round, currentLeader[0])
        leaderboard = []
        for prefix in players:
            testSpectrum = set(TheoreticalSpectrum(prefix))
            totalWeight = max(testSpectrum)
            score = len(spectrum & testSpectrum)/float(len(spectrum | testSpectrum))
            if (score > currentLeader[0]):
                currentLeader = [score, prefix]
            elif (score == currentLeader[0]):
                currentLeader += [prefix]
            if (totalWeight < target):
                leaderboard.append((score, prefix))
        remaining = len(leaderboard)
        if (remaining == 0):
            print "Done, no sequences can be extended"
            break
        leaderboard.sort(reverse=True)
        # Prune the larger of the top 5% or the top 5 players
        cut = leaderboard[max(min(5, remaining-1), int(remaining*cutThreshold))][0]
        players = [p+r for s, p in leaderboard if s >= cut for r in Daltons.iterkeys()]
        round += 1
    return currentLeader

spectrum = TheoreticalSpectrum(TyrocidineB1)
experimentalSpectrum = [mass for mass in spectrum if mass not in missingMass] + falseMass
%time winners = LeaderboardFindPeptide(experimentalSpectrum)
print winners
print len(winners) - 1, "Candidate residues with", winners[0], 'matches'
print TyrocidineB1, TyrocidineB1 in winners
```

```
400 Players in round 1 [0.0000]
480 Players in round 2 [0.0600]
1280 Players in round 3 [0.1200]
1560 Players in round 4 [0.2000]
2000 Players in round 5 [0.2745]
2600 Players in round 6 [0.3654]
3320 Players in round 7 [0.4615]
3520 Players in round 8 [0.5556]
3840 Players in round 9 [0.6545]
2400 Players in round 10 [0.8036]
160 Players in round 11 [0.8036]
Done, no sequences can be extended
CPU times: user 1.52 s, sys: 55 ms, total: 1.58 s
Wall time: 1.51 s
[0.8035714285714286, 'YQNFWPFLQV', 'YQNFWPFLKV', 'YQNFWPFIQV', 'YQNFWPFIKV', 'YKNFWPFLQV', 'YKNFWPFLKV', 'YKNFWPFIQV', 'YKNFWPFIKV', 'VQLFPWFNQY', 'VQLFPWFN
KY', 'VQIFPWFNQY', 'VQIFPWFNKY', 'VKLFPWFNQY', 'VKLFPWFNKY', 'VKIFPWFNQY', 'VKIFPWFNKY']
16 Candidate residues with 0.803571428571 matches
VKLFPWFNQY True
```

Next Time

- This method works well, but it relies on heuristics, and thus might miss the best answer
- Our methods still make a lot of simplifying assumptions
- Relying only on exact matches might mislead us
- We will continue to explore ways of assembling peptide sequences from a given experimental spectrum